#### REVIEW ARTICLE



# A systematic review and meta-analysis on the prevalence of antibiotic-resistant *Listeria* species in food, animal and human specimens in Iran

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**Abstract** The aim of this study was to determine the antimicrobial resistance characteristics of Listeria species isolated from foods and food processing environments, animal and human specimens in Iran. A systematic review of the papers published in Persian and English languages up to 20th May 2019 and indexed in the Scientific Information Database, PubMed, Scopus and Google Scholar databases using related keywords was conducted. Eligible articles were selected based on the predefined inclusion and exclusion criteria, followed by data extraction and meta-analysis using random-effects or fixed-effects models. A total of 27 articles were found reporting antibiotic resistance patterns of different Listeria species using disk diffusion method. Among Listeria species, Listeria monocytogenes resistance to commonly used antibiotics i.e. penicillin, ampicillin and gentamicin was as follows: 34.5%, 26.4%, 8.9% in isolates from foods and food processing environments, 47.1%, 29.5%, 9.2% in isolates from animal specimens and 56.8%, 29.5%, 32.4% in human strains, respectively. A high prevalence of L.

monocytogenes strains resistant to penicillin, ampicillin and gentamicin was observed in Iran. Our findings suggested that trimethoprim/sulfamethoxazole, vancomycin and ciprofloxacin can be used as alternatives in the treatment of human listeriosis in Iran due to their low resistance rates.

**Keywords** Antibiotic resistance · *Listeria* · Iran · Meta-analysis

#### Introduction

The genus Listeria consists of β-hemolytic, non-sporeforming, motile (at 22–28 °C; tumbling motility), Grampositive, facultative anaerobic rods which can be arranged as pairs or short chains (Carroll et al. 2016; Murray et al. 2015). Among 19 identified Listeria species, two species are considered as pathogens i.e. Listeria monocytogenes (L. monocytogenes) that regularly infects humans and Listeria ivanovii (L. ivanovii) that infects animals and rarely humans (Murray et al. 2015; Orsi and Wiedmann 2016). L. monocytogenes is a facultative intracellular food-borne pathogen that is widely found in nature and isolated from humans, wild and domestic animals, birds, insects, vegetation, water, food products and soil (Schuppler and Loessner 2010; Ramaswamy et al. 2007). Usually, the newborn (3.4 per 100,000), the elderly people (10 per 100,000), pregnant women (12 per 100,000) and patients with defects in cellular immunity are susceptible to Listeria infections which can be acquired via consumption of contaminated food products and mother to child transfer in utero or at birth (Murray et al. 2015; Olaimat et al. 2018). L. monocytogenes can cause neonatal diseases including early-onset listeriosis characterized by abortion, stillbirth,

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premature birth and high mortality rate which is acquired in utero. The pathogen can also cause late-onset listeriosis characterized by meningitis or meningoencephalitis with septicemia which is acquired 2-3 weeks after birth. The bacterium is also associated with a mild influenza-like illness and acute self-limited gastroenteritis in healthy adults, infections in pregnant women and meningitis in adults (Murray et al. 2015). In addition, the presence of Listeria species in food products is an important issue due to the consequent economic costs for the community in terms of consumer safety, and for the food industry (Ivanek et al. 2005). Hence, identification, control and treatment of Listeria infections are necessary. Combination regimens including a β-lactam alone or in combination with an aminoglycoside, i.e. either gentamicin with penicillin or gentamicin with ampicillin, are drugs of choice in the treatment of severe Listeria infections (Murray et al. 2015). Other antibiotics, such as trimethoprim-sulfamethoxazole, are successfully used to treat listeriosis but Listeria resistance to some of them has been reported, especially to fluoroquinolones, macrolides and tetracyclines (Murray et al. 2015; Olaimat et al. 2018). The prevalence of foodborne listeriosis varies between 0.1 and 11.3 cases per million persons in different countries with a mortality rate of  $\leq 30\%$  (Olaimat et al. 2018). In Iran, a systematic review and meta-analysis estimated the prevalence of L. monocytogenes to be 10%, 7% and 4% in humans, animals and food products, respectively (Ranjbar and Halaji 2018). However, no comprehensive information regarding antibiotic resistance of *Listeria* species in Iran has yet been presented. Therefore, the aim of this study was to determine the prevalence of antibiotic-resistant Listeria species, particularly L. monocytogenes, isolated from foods and food processing environments, animal and human specimens in Iran.

#### Methods

# Search strategy

According to the PRISMA guidelines (Liberati et al. 2009), the current meta-analysis was conducted on the prevalence of *Listeria* species antibiotic resistance isolated from foods and food processing environments, animal and human specimens in Iran. For this purpose, a systematic literature search was done to find published articles until May 20, 2019 through international databases such as PubMed, Scopus and Google Scholar, as well as an Iranian national database i.e. the Scientific Information Database (SID)

(https://www.sid.ir/En/Journal). We used three search terms including "antibiotic resistance", "Listeria" and "Iran" in both Persian and English languages. Finally, reference lists of the included eligible articles were checked for additional studies which might have been missed during databases searching.

#### Selection criteria

Identification of articles evaluating Listeria species antibiotic resistance was performed using the related keywords by two investigators independently. Retrieved articles were transferred to the EndNote reference management software. At first, the titles and abstracts and then the full texts were assessed based on inclusion and exclusion criteria. Studies with the following criteria were included in the meta-analysis: cross-sectional studies published in English or Persian languages, which were limited to Iran, Listeria species samples collected from foods and food processing environments, animal and human origins, and studies presenting data on the prevalence of antibiotic resistance. Non-original articles, duplicate reports, research evaluating drug resistance profiles only at the genus Listeria level or with insufficient data or small sample size and also articles measuring antibiotic resistance Listeria species isolated from mixed and non-specified samples were excluded.

#### **Quality assessment and data extraction**

The Joanna Briggs Institute (JBI) critical appraisal checklist was used to assess the quality of eligible studies (Munn et al. 2015). As shown, the main data were extracted and organized into Table 1 including the name of the first author, study publication date, *Listeria* species type, samples origins, the frequency of *Listeria* species, methods used for assessing susceptibility to different antibiotics and number of resistance to each antibiotic of *Listeria* species.

# Meta-analysis

The prevalence of *Listeria* species resistant to different antibiotics was estimated as percentage and 95% confidence intervals (CIs) through pooling the data using either random-effects model (when the heterogeneity was high among included studies) or fixed-effects model (when the heterogeneity was low). In this regard, heterogeneity was estimated using the  $I^2$  statistic and the Chi-square test with the Cochrane Q statistic (significant at  $I^2 > 25\%$  and



 Table 1
 Extracted information from eligible studies included in the meta-analysis

References	Listeria	Sample	Strain	Antibiotic		resistance (n)	(n)															
	sbecies	origin	(n)	PEN	AMP	GEN	TMP- SMX	TET	VAN	STR	CHL	CTX	ERY	CIP	СП 1	KAN	RIF ,	AMC (	CEF	CAZ	AMK	NAL
Maktabi et al. (2016)	L. innocua	Food	2	0	0	0	1	-	0	0	0	ND	0	QN	ND I	ND	ND	ND I	ND	ND	0	N Q
Rostami et al. (2015)	L. monocytogenes	Food	S	1	2	0	<u>R</u>	0	0	<u>R</u>	0	<u>R</u>	-	2	N ON	E E	N ON	E E	N Q	<u>R</u>	<u>R</u>	4
Rostami et al. (2015)	L. innocua	Food	7	-	0	0	S	0	0	N N	0	8	0	1	ND	E E	N ON	Q Q	N Q	<u>R</u>	S S	2
Rahimi et al. (2017)	L. monocytogenes	Food	S	S	8	_	Ð	5	2	N Q	_	N N	_	_	N ON	N D	E O	Q Q	N Q	N N	N Q	0
Rahimi et al. (2017)	L. innocua	Food	10	10	R	_	Q.	6	4	ND	2	N N	2	8	ND	N Q	N D	QZ QZ	ND ND	N N	N Q	0
Rahimi et al. (2017)	L. seeligeri	Food	2	2	8	0	<u>R</u>	-		<u>R</u>	0	<u>R</u>	0	0	ND	S S	N ON	e e	N Q	<u>R</u>	<u>R</u>	0
Rahimi et al. (2010)	L. monocytogenes	Food	19	9	5	-	S S	8	0	N Q	2	<u>R</u>	8	4	ND	E E	E O	Q Q	N Q	<u>R</u>	N Q	19
Rahimi et al. (2010)	L. innocua	Food	31	11	-	0	<u>R</u>	Ξ	0	N N	4	<u>R</u>	4	9	ND	E E	E O	E E	N Q	<u>R</u>	<u>R</u>	29
Rahimi et al. (2010)	L. seeligeri	Food	5	7	_	0	Q.	_	0	ND	_	N N	0	1	ND	E E	N D	QZ QZ	ND ND	N N	N Q	5
Rezai et al. (2018)	L. monocytogenes	Food	22	9	7	4	'n	14	N N	ND ND	7	R	6	15	5 I	Ę	N D	QZ QZ	ND	<u>R</u>	N N	N N
Rezai et al. (2018)	L. ivanovii	Food	16	S	∞	4	4	Ξ	ND	ND	4	N	5	9	3	Q Q	N D	QZ QZ	ND	N N	N Q	<u>R</u>
Rezai et al. (2018)	L. welshimeri	Food	6	8	7	0	8	2	N N	N N	3	N Q	3	7	2	E E	N D	Q Q	N N	N N	S S	N N
Rezai et al. (2018)	L. grayi	Food	12	S	9	2	5	7	ND ND	ND ND	2	ND	3	8	3 1	N Q	N ON	Q Q	N N	N N	N N	<u>R</u>
Rezai et al. (2018)	L. innocua	Food	18	6	4	9	ю	12	S	N N	4	N Q	4	Ś	4	Q Q	E C	Ę Ę	N Q	N N	N N	N
Rezai et al. (2018)	L. seeligeri	Food	6	8	8	2	7	2	<u>R</u>	N N	_	ND	4	2	1	2 Q	N D	Q Q	N N	N N	S S	<u>R</u>
Mojtahedi et al. (2004)	L. monocytogenes	Food	70	0	0	0	0	<u>R</u>	<u>R</u>	<u>R</u>	ND	Q.	5	S S	QN QN	2 Q	E E	Q Q	<del>S</del>	<u>R</u>	<u>R</u>	S
Mojtahedi et al. (2004)	L. innocua	Food	42	0	0	0	0	N N	N N	N N	ND	N N	N N	P.	ND	E E	N D	Q Q	N N	N N	S S	N N
Mojtahedi et al. (2004)	L. seeligeri	Food	∞	0	0	0	0	N N	ND ND	ND ND	ND	N N	ND ND	P.	ND	N Q	N D D	Q Q	N N	N N	N N	<u>R</u>
Kargar and Ghasemi (2011)	L. monocytogenes	Food	56	7	0	<u>S</u>	N Q	29	N Q	Q.	ND	<u>S</u>	12	<u>R</u>	ND ON	ND ND	ND ND	ND	QN Q	N Q	11	Ŋ



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References	Listeria species	Sample	Strain	Antibi	Antibiotic resistance (n)	ance (n)																
		ongin	(n)	PEN	AMP	GEN	TMP- SMX	TET	VAN	STR (	снг с	CTX	ERY	CIP (	CLI K	KAN R	RIF A	AMC C	CEF (	CAZ ,	AMK	NAL
Fallah et al. (2012)	L. ivanovii	Food	13	7	7	0	0	2	0	ND 3	S N	) QN	0	5 0		ND 0		ND N	ND N	ND	ND	ND
Fallah et al. (2012)	L. seeligeri	Food	7	0	0	0	0	_	0	ON ON	0		0	0 0		O QN		N ON	N ON	- ON	ND	<u>R</u>
Khalili Borujeni et al. (2013)	L. ivanovii	Food	'n	S	0	0	0	0	<u>R</u>	O Q	0	ON ON ON	0	Q Q	ND	Z Q	Z Q	QN QN	N Q	- E	NA	Ð
Rahimi et al. (2012b)	L. monocytogenes	Food	5	8	2	0	S S	8	0	ND 1	_	N ON	3	1	N QN	Z QZ	Z Q	N ON	S S	E E	ND	3
Rahimi et al. (2012b)	L. innocua	Food	14	7	0	0	Ð	4	0	ND 2	2 N	) QN	0	4	N ON	N ON	Z Q	N ON	N ON	- ON	N N	13
Abbasinejad et al. (2015)	L. monocytogenes	Food	8	0	N	0	N N	0	Ð	ND (N	0	ON ON	0	N N	ND	Z QZ	N ON	N ON	S S	- QN	ND	N N
Akrami- Mohajeri et al. (2018)	L. monocytogenes	Food	22	17	N N	4	0	19	0	10 1	N 71	8 8	∞	ND 5	. 7	-	6		S S		Q.	Ð
Akrami- Mohajeri et al. (2018)	L. innocua	Food	31	25	S	0	0	20	0	4	23 N	ND P	4	ND 2	0	0	-	2	P P	E E	<u>R</u>	Ð
Akrami- Mohajeri et al. (2018)	L. seeligeri	Food	10	4	S	0	0	5	0	5 3	2	S S	2	ON ON	0	-	-	_	N Q	- ON	N N	Ð
Soleymani Najaf Ababdi et al. (2017)	L. monocytogenes	Food	104	30	44	10	10	04	8	ND 2	25 N	e:	es.	20 8		ND 7		QN QN	N ON		Q.	Ð
Heidarzadeh et al. (2018)	L. monocytogenes	Human	22	10	10	∞	<u>R</u>	10	Q.	10 1	12 N	E D	10	4	ND	Z QZ	Z Q	N ON	QN QN		Q.	ND
Abdollahzadeh et al. (2016)	L. monocytogenes	Human	7	S	7	-	0	0	Q.	0	0 7		Ð	Q Q	ND	Z QZ	Z Q	N ON	N Q	- ON	N N	N Q
Kalani et al. (2015)	L. monocytogenes	Human	14	∞	0	N	<u>R</u>	2	Q.	0	4 Z	E.	3	0	N QN	Z QZ	Z Q	N ON	Z Q		QN N	NO
Lotfollahi et al. (2011)	L. monocytogenes	Human	6	7	0	N	<u>R</u>	0	Q.	1	7		0	0	N QN	N ON	Z Q	N ON	QN QN		Q.	ND
Dehkordi et al. (2013)	L. monocytogenes	Animal	227	66	29	77	<del>S</del>	162	<del>Q</del>	9 62	N 89	N OZ	75	27 N	N ON	Z QZ	N ON	S B	36 N	- ON	<u>Q</u>	N Q
Jamali et al. (2014)	L. monocytogenes	Animal	19	9	NO	0	<u>R</u>	11	0	ON ON	0	N N	7	ND 2	0	0	1		QZ QZ		Q.	N Q
Jamali et al. (2014)	L. innocua	Animal	5	0	ND QN	0	R R	-	0	ON ON	0	ND QN	0	O ON	0	0	0 (		Z QZ	<u> </u>	<u>R</u>	<u>R</u>



References	Listeria species Sample Strain Antibiotic	Sample	Strain	Antibio	tic resis	resistance (n)																
		origin	(u)	PEN	PEN AMP GEN	GEN	TMP- SMX	TET	VAN	STR	СНГ	CTX	CHL CTX ERY	CIP	CLI	KAN	RIF	CIP CLI KAN RIF AMC CEF	CEF	CAZ AMK	AMK	NAL
Jamali et al. (2014)	L. seeligeri	Animal	6	1 ND	N N	0	R	2	0	ND	0	N	0	N N	ND 0 0N	0	0	0	N Q	ND	N Q	ND
Jamali et al. (2014)	L. ivanovii	Animal	25	7	N	0	N N	4	0	N	0	N	S	R	4	0	0	3	S	N N	<u>R</u>	8
Jamali and Radmehr (2013)	L. monocytogenes	Animal	17	12	R	0	g	10	0	S	4	R	_	S	0	0	0	5	Ð	Ð	S	S
Jamali and Radmehr	L. innocua	Animal	κ	П	N	0	8	-	0	N	0	8	0	S	0	0	0	0	2	2	Ð	S

ERY erythromycin, CIP ciprofloxacin, CLI clindamycin, KAN kanamycin, RIF rifampin, AMC amoxicillin-clavulanic acid, CEF cephalothin, CAZ ceftazidime, AMK amikacin, NAL nalidixic PEN penicillin, AMP ampicillin, GEN gentamicin, TMP/SMX trimethoprim/sulfamethoxazole, TET tetracycline, VAN vancomycin, STR streptomycin, CHL chloramphenicol, CTX cefotaxime, acid, ND not determined

p < 0.1). Asymmetry of funnel plots was explored as an indicator of potential publication bias. Meta-analysis was performed using the Comprehensive Meta-Analysis (CMA) software version 2.2 (Biostat, Englewood, NJ).

#### Results

# Literature search and study characteristics

Of 1069 collected reports after searching in national and international databases, a total of 27 articles met predefined inclusion and exclusion criteria and were found to be eligible for the analysis of Listeria species antibiotic resistance (Fig. 1). Included studies were reported from Ahvaz, Bandar Torkaman, Chahar Mahal & Bakhtiyari, Isfahan, Karaj, Kurdistan, Lorestan, Marvdasht, Mashhad, Mazandaran, Shiraz, Tehran, Urmia and Yazd. As shown in Table 1, 20, 4 and 3 the research papers assessed antibiotic resistance profiles of Listeria species isolated from foods and food processing environments, human and animal origins, respectively. Food products as raw, ready-to-cook and ready-to-eat included seafood products, milk and traditional dairy products, processed meat, poultry products as well as market and processing environments which have been used for food products. Clinical specimens were collected from pregnant women with vaginitis (vaginal swab) and woman with spontaneous abortion (blood, urine, placental tissue, fecal and vaginal swabs). Also, animal specimens were obtained from ducks, geese, bovine, ovine, caprine, buffalo and camel species. L. monocytogenes was the most common Listeria species isolated from foods and food processing environments (57.9% samples), human (100% samples) and animal (86.2% samples) specimens.

# Characteristics of *Listeria* species antibiotic resistance in Iran

All eligible studies used disk diffusion technique to determine antimicrobial susceptibility of *Listeria* species isolated in Iran. The results of antibiotic resistance according to the type of sample are shown in Table 2 for *L. monocytogenes*, *L. ivanovii*, *L. innocua*, *L. seeligeri*, *L. grayi* and *L. welshimeri*. As shown in Table 2, there is a heterogeneity among some studies in determining the prevalence of antibiotic resistance in *Listeria* species. In this case, random-effects model was used for pooling the data. Also, funnel plots were asymmetric suggesting potential publication bias in estimating the percentage of resistance for some antibiotics (Fig. 2).



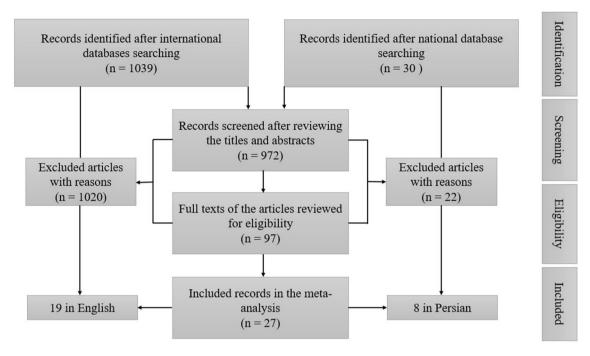


Fig. 1 Flowchart of literature search and study selection for meta-analysis

# **Discussion**

Listeria together with other pathogens including Campylobacter, Vibrio, Salmonella, Shigella, Escherichia, Yersinia, Staphylococcus, Clostridium and Bacillus are causative agent for more than 90% of food poisonings (Jalalpour 2012). It has been suggested that antibiotic therapy is necessary in severe cases of infection but drug resistance has increased among food-borne pathogens during the past few decades. Drug-resistant microbial species are a significant threat to public health, food security and treatment of diseases, and account for longterm hospitalization, increased mortality rates and enormous medical costs in the world (Olaimat et al. 2018; Jamali et al. 2014). One of the main food-borne microbes associated with major public health concern, especially in the food industry, is L. monocytogenes (Olaimat et al. 2018). In the present study, resistance rates of L. monocytogenes as well as other *Listeria* species in food products were high against some antibiotics (Table 1). Antibiotic resistance of food-borne L. monocytogenes pathogen may be either intrinsic or acquired due to genetic alterations or adaptation to environmental stresses including physical (e.g., heat and high pressure), chemical (e.g., acids and salts) and biological (e.g., microbial antagonism) stressors during food production and in the food processing environments (Olaimat et al. 2018; Lungu et al. 2011). Studies have also shown that factors such as the use of antimicrobial compounds in food preservation (via inducing efflux pumps), as well as sublethal exposure to disinfectants in food processing environments may contribute to the emergence of antibiotic-resistant food isolates of L. monocytogenes (Olaimat et al. 2018). Since L. monocytogenes is isolated from ubiquitous sources, the food chain can play an important role in transferring antibiotic resistant strains between animals and humans (Olaimat et al. 2018). In this study, antimicrobial resistance of L. monocytogenes isolated from human specimens against antibiotics used in primary therapy of Listeria infections i.e. penicillin, ampicillin and gentamicin was found to be 56.8%, 29.5% and 32.4% (Fig. 3), respectively. These prevalence rates are higher than the results of L. monocytogenes isolated from food (34.5%, 26.4% and 8.9%) and animal (47.1%, 29.5% and 9.2%) origin specimens. Frequent use of these antibiotics as the first-line drugs in listeriosis treatment as well as acquisition of resistance genes from the commensal unrelated bacterial species in foods and food processing environments through mobile genetic elements and efflux pumps can justify the high levels of L. monocytogenes resistance in human samples (Olaimat et al.



Table 2 Characteristics of Listeria species antibiotic resistance isolated in Iran

Listeria species	Sample origin		Antibiotic resistance (n)	sistance (n)								
			PEN	AMP	GEN	TMP-SMX		VAN	STR	CHL	CTX	ERY
L. monocytogenes	Food	%	34.5	26.4	8.9	5.9	40.5	5.8	22	17.7	29.2	17.6
		95% CI	24.4-46.2	16.2–39.8	5.3-14.6	2.6–12.8		2.5-13.1	9-44.6	9.7–30.3	8.66-0	10.2–28.6
		$I^2$	81.9	83.3	54.6	9.69		69	69.4	83.9	91.9	83.3
		õ	8.66	84	35.2	23		35.5	13	2.66	12.3	101.9
		d	0.00	0.00	0.00	0.00		0.00	0.01	0.00	0.00	0.00
	Human	%	56.8	29.5	32.4	6.3		ı	28.7	27.8	83.1	8.97
		95% CI	42.7–69.9	4.3–79.6	17.6–51.7	0.4–53.9		1	9.7–60.1	10.6-55.8	55.3-95.1	9.4–56.2
		$I^2$	6.7	75.2	10.2	0		ı	61.5	60.1	0	57.3
		õ	3.2	12.1	1.1	0		1	7.7	7.5	0.76	4.6
		d	0.36	0.00	0.29	1.0		1	0.05	0.05	0.38	0.09
	Animal	%	47.1	29.5	9.2	I		2.6	34.8	23.3	ı	16.5
		95% CI	30.3-64.6	23.9–35.8	0.9 - 52.1	ı		0.4–16.5	28.9-41.2	11.2-42.2	ı	4.8-43.4
		$I^2$	64	0	75.8	ı		0	0	51	ı	72.6
		õ	5.5	0	8.2	I		0	0	4	ı	7.3
		d	90.0	1.0	0.01	ı		0.95	1.0	0.13	ı	0.02
L. ivanovii	Food	%	41.5	29.5	10.5	14.4		8	15.8	20.9	2.5	12
		95% CI	19.9–67	9.6–62.4	3-30.9	6.5-28.9		0.4 - 18.4	5.2-39.2	8.1-44.4	0.2-29.8	3.3-34.9
		$I^2$	60.7	64.9	31.8	21.9		0	0	42.2	0	45.3
		õ	7.6	8.5	4.4	3.8		0	0	5.1	0	5.4
		р	0.05	0.03	0.22	0.27		0.85	1.0	0.15	1.0	0.13
	Human	%	I	I	1	I		ı	ı	I	I	1
		95% CI	ı	ı	ı	ı		1	ı	ı	I	ı
		$I^2$	ı	ı	ı	ı		ı	ı	1	ı	I
		õ	I	1	I	ı		ı	ı	1	ı	I
		b	I	I	I	I		ı	1	I	ı	I
	Animal	%	28		1.9	ı		1.9	I	1.9	ı	20
		95% CI	14-48.2		0.1–24.4	ı		0.1-24.4	I	0.1–24.4	I	8.6-40
		$I^2$	0		0	1		0	ı	0	ı	0
		õ	0		0	1		0	I	0	I	0
		d	1.0	1	1.0	I		1.0	I	1.0	ı	1.0
L. innocua	Food	%	43.7	12	4.8	6		4.7	15.5	15.3	1	12.4
		95% CI	30.3–58.1	3.9–31.4	1.9–11.5	4.1–18.3		1.5-13.2	9.1–25.2	6.8–30.9	0.1-14.6	9.3–16.5
		$I^2$	79.3	88.1	58.4	40.6		2.09	0	83.5	0	0
		õ	53.3	67.7	26.4	8.4		22.9	0.2	2.09	0	9.2
		d	0.00	0.00	0.00	0.13		0.00	0.88	0.00	1.0	0.51



Table 2 continued

Listeria species	Sample origin		Antibiotic re	sistance (n)								
			PEN AMP	AMP	GEN	TMP-SMX	TET	VAN	STR	CHL	CTX	ERY
	Human	%	ı	I		ı	I			ı	I	ı
		95% CI	1	1		1	ı			ı	ı	ı
		$I^2$	1	1		ı	ı			ı	ı	ı
		õ	I	I		ı	I			I	ı	ı
		d	1	I		ı	I			ı	I	I
	Animal	%	20	ı		ı	25.5			10.2	1	10.2
		95% CI	3.8–61.3	ı		I	6.4-63.3			1.4-47.3	ı	1.4-47.3
		$I^2$	0	ı		ı	0			0	ı	0
		õ	0.7	ı		ı	0.1			0	ı	0
		d	0.37	1		I	0.67			0.83	1	0.83
L. seeligeri	Food	%	26.1	14.7		12.1	30.9			15	2	18
		95% CI	13.9–43.7	7.2–27.7		5.7–23.9	20.4-43.7			7.6-27.7	0.1 - 25.1	10.1–29.9
		$I^2$	36.4	0		0	8			0	0	0
		õ	12.5	4.2		1.9	7.6			4.7	0	6.2
		d	0.12	0.50		0.74	0.36			89.0	1.0	0.50
	Human	%	1	ı		ı	1			ı	ı	ı
		95% CI	1	I		I	ı			ı	I	ı
		$I^2$	I	I		I	I			I	I	I
		õ	I	I		ı	ı			ı	ı	ı
		d	I	ı		ı	ı			ı	I	1
	Animal	%	11.1	I		I	22.2			s.	ı	w
		95% CI	1.5–50	ı		I	5.6-57.9			0.3-47.5	1	0.3-47.5
		$I^2$	0	ı		ı	0			0	1	0
		õ	0	I		I	0			0	ı	0
		d	1.0	1		ı	1.0			1.0	ı	1.0
L. grayi	Food	%	37.9	45.1		41.7	47.3			16.7	ı	23.5
		95% CI	17.2–64.1	22.2–70.4		18.5–69.2	15.3-81.7			4.9-43.7	I	8.5-50.5
		$I^2$	0	0		0	27.5			0	ı	0
		$\tilde{o}$	0.5	6.0		0	1.3			0	I	0
		d	0.44	0.33		1.0	0.24			1.0	I	0.76
	Human	%	I	I		I	I			I	I	I
		95% CI	I	I		ı	ı			ı	ı	ı
		$I^2$	I	I		ı	1			ı	1	1
		$\tilde{o}$	1	ı		I	1			ı	ı	ı
		р	1	ı		1	1			1	1	1



21.7-94.5 2.9-47.1 ERY NAL CTX12.4-54.9 AMK 0.03 4.4 2.9-47.1 CHL CAZ4 STR 15.7 CEF20.5–56.4 AMC 0.48 0.4 12.3 2.2 0.31 TET TMP-SMX 11.1-66.7 20.6 0.00 RIF 1.8 - 12KAN GEN Antibiotic resistance (n)
CIP CLI **14.2** 6–30 50.1 0.00 88 Antibiotic resistance (n) AMP 16.3-37.4 5.6-28.4 13.3 20.4-54.7 0.00 PEN 95% CI 95% CI 95% CI 95% CI 95% CI 95% CI Sample origin Sample origin Human Food Animal Human Animal Food L. monocytogenes Listeria species Listeria species L. welshimeri



Table 2 continued

Table 2 continued

NAL AMK 10.3 6.3–16.4 23.6 3.9 0.26 Antibiotic resistance (n)
CIP CLI Sample origin Animal Listeria species L. ivanovii



Listeria species	Sample origin		Antibiotic resistance (n)	istance (n)							
			CIP	CLI	KAN	RIF	AMC	CEF	CAZ	AMK	NAL
L. seeligeri	Food	%	16.1	7.3	4.6	8.1	11.9	14	2	ı	60.4
		95% CI	6.5–34.8	2.1–22.4	1.2–16.8	3.1–19.7	3–37.3	5.3–31.9	0.1–25.1	I	2.9–98.7
		$I^2$	0	0	0	0	0	0	0	ı	71.4
		õ	1	0.33	0.5	0.1	0	0.1	0	I	3.5
		d	0.89	0.95	0.89	86.0	0.78	99.0	1.0	I	90.0
	Human	%	ı	I	I	1	I	I	ı	ı	1
		95% CI	I	I	I	I	I	I	I	I	I
		$I^2$	I	I	I	I	I	I	I	I	ı
		õ	ı	I	ı	1	I	I	ı	ı	ı
		d	I	I	I	I	I	I	I	I	I
	Animal	%	I	w	w	w	w	I	ı	I	ı
		95% CI	ı	0.3-47.5	0.3-47.5	0.3-47.5	0.3-47.5	I	1	I	ı
		$I^2$	ı	0	0	0	0	I	ı	I	ı
		õ	I	0	0	0	0	I	I	I	ı
		d	ı	1.0	1.0	1.0	1.0	I	I	I	1
L. grayi	Food	%	23.5	25	16.7	1	I	16.7	1	1	1
		95% CI	8.5–50.5	8.3–55.2	1 - 80.6	I	I	1–80.6	I	I	ı
		$I^2$	0	0	0	ı	I	0	I	I	1
		0	0	0	0	I	I	0	ı	I	1
		d	0.76	1.0	1.0	I	I	1.0	ı	I	I
	Human	%	ı	I	I	ı	I	I	I	I	1
		95% CI	ı	I	I	I	I	I	ı	I	1
		$I^2$	I	I	I	I	I	I	I	I	ı
		õ	I	I	I	I	1	I	1	1	ı
		b	ı	ı	I	ı	1	I	ı	ı	ı
	Animal	%	ı	ı	I	ı	1	I	ı	ı	ı
		95% CI	I	I	I	I	1	I	I	1	1
		$I^2$	I	I	I	I	I	I	I	I	1
		Õ	1	I	I	I	I	I	1	ı	ı
		b	ı	I	I	ı	ı	I	1	ı	I
L. welshimeri	Food	%	22.2	15.8	4.8	4.5	4.5	22.2	I	I	1
		95% CI	8.6-46.5	4.5–42.6	0.7–27.2	0.3-44.8	0.3-44.8	5.6–57.9	I	ı	1
		$I^2$	0	14.7	0	0	0	0	I	I	1
		õ	0	1.1	0	0	0	0	1	ı	ı
		d	1.0	0.27	96.0	1.0	1.0	1.0	1	ı	1



Table 2 continued

	A
	Sample origin
Table 2 continued	Listeria species S

Listeria species	Sample origin		Antibiotic	resistance (n)	Antibiotic resistance (n)						
			CIP	CLI	KAN	RIF	CLI KAN RIF AMC	CEF	CAZ	CAZ AMK NA	NAL
	Human	%	ı	ı	1	ı	ı	ı	ı	ı	
		95% CI	ı	ı	ı	ı	ı	ı	ı	1	I
		$I^2$	ı	I	ı	ı	I	I	ı	I	ı
		õ	1	ı	ı	I	1	ı	I	1	1
		d	I	I	I	I	I	I	I	I	I
	Animal	%	I	I	I	I	I	ı	ı	ı	I
		95% CI	ı	ı	I	I	I	ı	ı	I	ı
		$I^2$	ı	I	I	I	I	I	I	I	I
		Õ	I	ı	ı	I	I	ı	I	I	I
		D	ı	I	I	I	I	ı	ı	I	ı

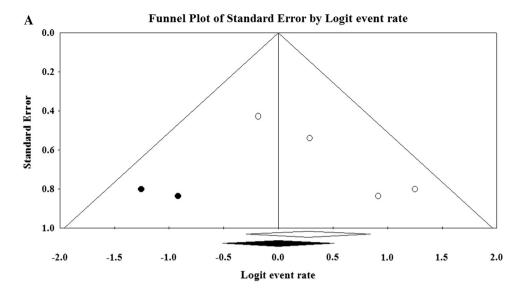
ERY erythromycin, CIP ciprofloxacin, CLI clindamycin, KAN kanamycin, RIF rifampin, AMC amoxicillin-clavulanic acid, CEF cephalothin, CAZ ceftazidime, AMK amikacin, NAL nalidixic PEN penicillin, AMP ampicillin, GEN gentamicin, TMP/SMX trimethoprim/sulfamethoxazole, TET tetracycline, VAN vancomycin, STR streptomycin, CHL chloramphenicol, CTX cefotaxime,

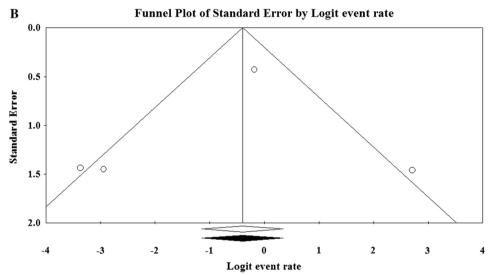
30ld indicates total resistance rate to each antibiotic

2018; Abdollahzadeh et al. 2016). Additionally, in penicillin-allergic individuals, vancomycin or trimethoprim/sulfamethoxazole are recommended for the treatment of invasive listeriosis (Abdollahzadeh et al. 2016). The prevalence of vancomycinand trimethoprim/sulfamethoxazole-resistant L. monocytogenes isolated from different origins was as follow: 5.9% to trimethoprim/sulfamethoxazole and 5.8% to vancomycin in food specimens. 6.3% to trimethoprim/sulfamethoxazole in human specimens and 2.6% to vancomycin in animal specimens (Table 2). The antibiotic resistance of L. monocytogenes isolated from food products in different countries was as follows: 100% to penicillin, 4.8% to gentamicin, 57.1% to vancomycin and 4.8% to trimethoprim/sulfamethoxazole in India, 93.8% to penicillin, 3.1% to gentamicin, 46.8% to vancomycin and 6.3% to trimethoprim/sulfamethoxazole in Egypt, and 100% to penicillin, 7.8% to gentamicin, 45.1% to vancomycin and 11.7% to trimethoprim/sulfamethoxazole in Yemen (Obaidat et al. 2015). Another study from Turkey showed 66.7% resistance to penicillin, 75% to ampicillin, 8.3% to gentamicin, 0% to vancomycin and 50% to trimethoprim/sulfamethoxazole (Aras and Ardic 2015). Also, Ha et al. in a study in South Korea indicated that ampicillin, gentamicin, vancomycin and trimethoprim/sulfamethoxazole resistance rates of L. monocytogenes isolated from food products were 97%, 0%, 0% and 0%, respectively, (Ha et al. 2017). On the other hand, it is necessary to monitor the extensive use of antibiotics as supplements to enhance the growth or prevent, control and treat diseases in animals or as pesticide on fruits, and also use of sub-inhibitory concentrations of antibiotics in human diseases treatment which can lead to the emergence of the antibiotic resistance and effect on human infection therapy (Olaimat et al. 2018). It is worth noting that tetracycline is frequently used for treating animal infections in Iran (Akrami-Mohajeri et al. 2018). This drug is also used in treating human infections (Olaimat et al. 2018). Our results indicated that L. monocytogenes strains isolated from food products (40.5%), animal (69.5%) and human (18.3%) samples exhibited a high resistance rate to tetracycline. Similar results were observed for other Listeria species (Table 2). However, 3% and 6% of L. monocytogenes isolates from food products in Italy and Canada were resistant to tetracycline, respectively (Abdollahzadeh et al. 2016). In pregnant women, erythromycin is an antibiotic of choice to treat listeriosis (Olaimat et al. 2018). However, the current study showed that 26.8% of human isolates of L. monocytogenes were resistant to erythromycin. Other antibiotics used to treat listeriosis are rifampicin, chloramphenicol and fluoroquinolones (Olaimat et al. 2018). Among them, chloramphenicol-resistant L. monocytogenes strains isolated from food (17.7%), animal (27.8%) and human (23.3%) origins and



Fig. 2 Funnel plots depicting publication bias of studies reporting the prevalence antibiotic resistance of *L. monocytogenes* to penicillin (a) and ampicillin (b) in human specimens





ciprofloxacin-resistant *L. monocytogenes* strains isolated from food origin (25.4%) showed high resistance rates. However, rifampin-resistant *L. monocytogenes* strains isolated from food (5.4%) and animal (2.6%) origins had low resistance rates. In addition to the pathogen source, other species of the genus *Listeria* can affect the emergence of *L. monocytogenes* antibiotic resistance in through genetic material exchange (Allen et al. 2016). Therefore, investigation of the antibiotic resistance of other *Listeria* species is also important. Our study showed that resistance of *Listeria* species isolated from foods and food processing environments and animal sources in Iran was high to some antibiotics (Table 2).

# Conclusion

The present systematic review and meta-analysis warns about the emergence of *Listeria* species resistant to different antibiotics in different pathogen sources in Iran. For example, a high prevalence of *L. monocytogenes* strains resistant to commonly used antibiotics in treating human listeriosis i.e. penicillin, ampicillin and gentamicin was observed in Iran. In addition, other species of the genus *Listeria* indicated high resistance rates to some antibiotics. Therefore, to reduce the spread of *Listeria* drug resistance in Iran, excessive use of antibiotics either in the treatment of human infections or in animal feeding as growth supplements and prophylaxis should be controlled and limited. Nonetheless, our findings suggest that trimethoprim/sulfamethoxazole, vancomycin and ciprofloxacin can be



Fig. 3 Forest plots depicting the prevalence of antibiotic resistance of *L. monocytogenes* to penicillin (a), ampicillin (b) and gentamicin (c) in human specimens

#### $\mathbf{A}$

Study name		Statist	ics for e	ach study				Event r	ate and	95% CI	
	Event rate	Lower limit		Z-Value	p-Value	Total					
Heidarzadeh	0.455	0.265	0.659	-0.426	0.670	10/22					
Abdollahzadeh	0.714	0.327	0.928	1.095	0.273	5/7				-	┣┤
Kalani	0.571	0.316	0.794	0.533	0.594	8 / 14				-	-
Lotfollahi	0.778	0.421	0.944	1.562	0.118	7/9					_
	0.568	0.427	0.699	0.943	0.346					•	
							-1.00	-0.50	0.00	0.50	1.00
							1	Favours A		Favours I	2

Meta Analysis

#### В

Study name		Statist	ics for e	ach study				Event ra	ate and	95% CI	
	Event rate	Lower limit		Z-Value	p-Value	Total					
Heidarzadeh	0.455	0.265	0.659	-0.426	0.670	10/22					
Abdollahzadeh	0.938	0.461	0.996	1.854	0.064	7/7				-	
Kalani	0.033	0.002	0.366	-2.341	0.019	0 / 14				_	
Lotfollahi	0.050	0.003	0.475	-2.029	0.042	0/9					
	0.295	0.043	0.796	-0.765	0.444				<b>-</b>		-
							-1.00	-0.50	0.00	0.50	1.00
								Favours A		Favours l	В

Meta Analysis

# $\mathbf{C}$

Study name		Statist	ics for e	ach study				Event r	ate and	<u>95% C</u> I	
	Event rate	Lower limit		Z-Value	p-Value	Total					
Heidarzadeh	0.364	0.193	0.577	-1.263	0.207	8 / 22			-		
Abdollahzadeh	0.143	0.020	0.581	-1.659	0.097	1 / 7				$\rightarrow$	
	0.324	0.176	0.517	-1.798	0.072				•		
							-1.00	-0.50	0.00	0.50	1.00
							F	avours 2	A F	avours l	В

Meta Analysis

alterative antibiotics, due to their low resistance rates, in the treatment of human listeriosis in Iran. Finally, we recommend evaluating mechanisms of antibiotic resistance in *Listeria* species, especially *L. monocytogenes* in Iran.

#### Compliance with ethical standards

Conflict of interest The authors declare that there is no conflict of interest.

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